# Why don't all whales have cancer? A novel hypothesis resolving Peto's paradox

John D. Nagy,<sup>1,\*,†</sup> Erin M. Victor,\* and Jenese H. Cropper\*

\*Department of Life Sciences, Scottsdale Community College, 9000 E. Chaparral Rd., Scottsdale, AZ 85256, USA; \*Department of Mathematics and Statistics, Arizona State University, Tempe, AZ 85287-1804, USA

**Synopsis** Larger organisms have more potentially carcinogenic cells, tend to live longer and require more ontogenic cell divisions. Therefore, intuitively one might expect cancer incidence to scale with body size. Evidence from mammals, however, suggests that the cancer risk does not correlate with body size. This observation defines "Peto's paradox." Here, we propose a novel hypothesis to resolve Peto's paradox. We suggest that malignant tumors are disadvantaged in larger hosts. In particular, we hypothesize that natural selection acting on competing phenotypes among the cancer cell population will tend to favor aggressive "cheaters" that then grow as a tumor on their parent tumor, creating a hypertumor that damages or destroys the original neoplasm. In larger organisms, tumors need more time to reach lethal size, so hypertumors have more time to evolve. So, in large organisms, cancer may be more common and less lethal. We illustrate this hypothesis *in silico* using a previously published hypertumor model. Results from the model predict that malignant neoplasms in larger organisms should be disproportionately necrotic, aggressive, and vascularized than deadly tumors in small mammals. These predictions may serve as the basis on which to test the hypothesis, but to our knowledge, no one has yet performed a systematic investigation of comparative necrosis, histopathology, or vascularization among mammalian cancers.

#### Introduction

Cancer afflicts most, perhaps all, classes of vertebrates and appears to be most common in mammals (Galis 1999; Galis and Metz 2003). Basic cancer-causing mechanisms are similar among mammalian species, and many tumor-suppressing systems are conserved (Leroi et al. 2003), which allows us to model the disease in humans by using rodents and other mammals. Nevertheless, malignant neoplasia is somehow fundamentally different in different mammalian species, a fact recognized over 30 years ago by Peto et al. (1975). This fundamental difference is implied by what is now called "Peto's paradox" (Leroi et al. 2003).

The paradox begins with the following logical argument. If all mammalian cells were roughly equally susceptible to oncogenic mutations, had equivalent tumor-suppressing systems, and neoplastic cells were equally viable in all mammals, then the number of cells susceptible to malignant transformation in a given organism would be roughly proportional to its body size, and the probability that a given susceptible cell transitions to malignancy would be an increasing function of time, as observed in humans (Cole and Rodu 2001). Therefore, one would anticipate that cancer incidence would correlate positively, at least roughly, with the product of body mass and longevity. Within a species, this variation could be trivial, but among mammalian species, which vary in mass from  $2 \times 10^{-3}$  kg to 190,000 kg—a small bumblebee bat, *Craseonycteris thonglongyai* versus a large blue whale, *Balaenoptera musculus* (Pereira et al. 2006)—these correlations should be marked enough to measure, again under the assumption that cancer etiology and pathogenesis are roughly consistent among all mammals.

A review of the literature, however (discussed subsequently), reveals no obvious correlation between cancer incidence and body mass, in agreement with earlier assessments (Leroi et al. 2003). In general, cancer is easy to find in most mammals, but populations in which cancer is a leading cause of death seem to be special cases associated with an external etiologic agent and show no obvious association with body size, at least not

From the symposium "Ecology and Evolution of Disease Dynamics" presented at the annual meeting of the Society for Integrative and Comparative Biology, January 3–7, 2007, Phoenix, Arizona.

<sup>&</sup>lt;sup>1</sup>E-mail: john.nagy@sccmail.maricopa.edu

Integrative and Comparative Biology, volume 47, number 2, pp. 317-328

doi:10.1093/icb/icm062

Advanced Access publication June 28, 2007

<sup>©</sup> The Author 2007. Published by Oxford University Press on behalf of the Society for Integrative and Comparative Biology. All rights reserved. For permissions please email: journals.permissions@oxfordjournals.org.

what one would expect given masses that range over eight orders of magnitude. Therefore, we conclude that cancer etiology and pathogenesis are not consistent across the class. In other words, we conclude that Peto's is a legitimate paradox, the resolution to which is neither obvious nor trivial.

Here we suggest a hypothesis that could resolve Peto's paradox. In particular, we hypothesize that malignant cell populations in larger organisms are more susceptible to invasion by selfish "cheater" phenotypes, and these selfish cells damage or destroy the tumor from within, much like the tumor damages or destroys its host from within. In essence, the cheater population forms a tumor-within-atumor, or "hypertumor" (Nagy 2004). We use mathematical models and computer simulations to illustrate the hypertumor-mechanism hypothesis and generate practically testable predictions.

# Cancer epidemiology in mammals and Peto's paradox

Although current research is unable to establish ubiquity of cancer among mammals, evidence points in that direction. In particular, malignant neoplasia is widely reported in domestic animals and wildlife, both free-ranging and captive (Table 1). Beyond this, however, surprisingly little is known.

#### Cancer in wildlife

The majority of studies on cancer in wild mammals are case reports of individuals in zoos or sporadic cases observed in the wild (see references in Table 1). Few solid epidemiological studies in wild populations exist, largely because size of the population at risk is either unknown or ill-defined (Martineau et al. 2002). What evidence does exist suggests that incidence of malignant neoplasia in wild mammals tends to remain well below that in humans. For example, among captive wildlife, crude cancer risk (number of cancers in animals that die of natural causes) varies between about 1.5% and 4.5% (Galis and Metz 2003). In a population of Swedish roe deer, crude risk of all neoplasia is only 2% (Aguirre et al. 1999). In comparison, 20-30% of people in most countries suffer cancer at some time in their lives (Bishop 1989; Muirhead et al. 2004).

In a few cases, cancer in wild populations approaches or even exceeds that in humans, but in each case the cause is a well-defined, nongeneric agent. For example, the entire population of Tasmanian devils, a marsupial carnivore of Tasmanian Australia, are threatened by one particular cancer, called devil facial tumor disease (DFTD). Oddly, this cancer behaves as an emerging infectious disease. Scant evidence of DFTD existed prior to 1995, but by 1996 it had spread across half of the devil's range (Hawkins et al. 2006). In some areas, local prevalence approaches or even exceeds 80%, and population density appears to correlate negatively with prevalence (Hawkins et al. 2006). The disease is characterized by massive, infiltrating facial tumors that eventually impede the animal's ability to feed (Pearse and Swift 2006). Histologically, they appear to be sarcomas of neuroectodermal origin (Hawkins et al. 2006). Interestingly, karyotypes of cells from tumors in 11 different devils all show precisely the same complex aneuploidy, suggesting that the tumors pass as allografts from animal to animal during combat (Pearse and Swift 2006). In essence, this cancer metastasizes among, not just within, hosts.

Woodchucks also suffer high cancer rates, especially in the liver, but again the cause is a specific etiological agent. Some woodchuck populations harbor a hepatitis virus that, as in humans, causes a variety of hepatic diseases, including hepatocellular carcinoma (Snyder et al. 1982; Roth et al. 1985; Menne and Cote 2007). In fact, woodchucks have become an important animal model for hepatitis virus infection in humans (Menne and Cote 2007), so most literature focuses on experimentally induced infection. In one study of natural infection, however, 13 of 16 (81%) infected woodchucks also suffered hepatocellular carcinoma, with one distant metastasis, compared to one liver adenoma in 149 (0.7%) virus-free animals (Roth et al. 1985). Another study of uninfected woodchucks uncovered only two neoplasms in 147 animals (1.4%) (Roth et al. 1991). Other solid neoplasms reported in woodchucks include fibrosarcoma (Young and Webster 1985), teratoma, seminoma (Anderson and Johnson 1988), meningioma (Podell et al. 1988), pleural mesothelioma (Kang et al. 2004), and leiomyosarcoma (Kang et al. 2005), although these are all sporadic cases.

Perhaps the best nonhuman cancer epidemiological work has been performed on cetaceans—whales, porpoises, and dolphins. The disease has been identified in many cetacean species (Table 1). Most of these examples are sporadic case reports typical in wildlife cancer literature, but in one population of belugas inhabiting the St. Lawrence estuary, the incidence of cancer approaches or perhaps even exceeds that of humans (Martineau et al. 2002). The cause appears to be agricultural and industrial pollution of the estuary (De Guise et al. 1994, 1995; Martineau et al. 1994, 2002; Muir et al. 1996; Letcher et al. 2000). Common name

Virginia opossum

Tasmanian devil

Domestic dog

Island gray fox

Asian golden cat

House cat

Bengal tiger

African lion

Snow leopard

Domestic ferret

Cottontail Rabbit

House mouse

Woodchuck

Coypus

Rat

White-tailed Jackrabbit

White-footed mouse

California sea lion

African hedgehog

Domestic horse

Black rhinoceros

Northern elephant seal

Meerkat

Binturong

Coyote

Red fox

Table 1 A partial list of reported cancer in mammals and dinosaurs

#### Scientific name Reference Didelphis virginiana Prater et al. (1999) Sarcophilus laniarius Pearse and Swift (2006) Canis familiaris Michell (1999) Canis latrans Bekoff and Gese (2003) Vulpes vulpes Hirayama et al. (1999) Roemer et al. Urocyon littoralis (2004)Felis domesticus Mayr et al. (2000)Catopuma temminckii Rao and Acharjyo(1985) Panthera tigris Powe et al. (2005) Panthera leo Sakai et al. (2003)Uncia uncia Murata et al. (2003) Mustela putorius furo Jones et al.

Suricata suricata

Arctictis binturong

Sylvilagus spp.

Lepus townsendii

Mus musculus

Rattus spp.

Peromyscus leucopus

Marmota monax

Myocastor coypus

Zalophus californianus

Mirounga angustirostris

Atelerix albiventris

Equus caballus

Diceros bicornis

(2006)

Singh et al. (2005)

Klaphake et al. (2005)

Syverton et al. (1950)

Jardine et al. (2004)

Hirst and Balmain (2004)

Russo and Russo (1996)

Parnell et al. (2005)

Menne and Cote (2007)

Keymer et al. (1999)

Fauguier et al.

Raymond and

White (1999)

Plummer et al.

Radcliffe (2000)

Acevedo-Whitehouse et al.

(1999)

(2003)

(2007)

Paglia and

#### Table 1 Continued

Common name	Scientific name	Reference	
Asian elephant	Elaphis maximus	Liu et al. (2004)	
Dromedary camel	Camelus dromedarius	Vitovec (1982)	
Pig	Sus scrofa	Kleinschmidt et al. (2006)	
Western roe deer	Capreolus capreolus	Aguirre et al. (1999)	
Pere David's deer	Elaphurus davidianus	Yoon et al. (1999)	
Nubian ibex	Capra nubiana	Wooldridge et al. (1999)	
Domestic goat	Capra hircus	Whitney et al. (2000)	
Cattle	Bos taurus	Campo (1997)	
Domestic sheep	Ovis aries	Palmarini et al. (1999)	
Bottlenose dolphin	Tursiops truncatus	Martineau et al. (2002)	
Atlantic spotted dolphin	Stenella frontalis	Martineau et al. (2002)	
Pantropical spotted dolphin	Stenella attenuata	Martineau et al. (2002)	
Pacific white sided dolphin	Lagenorhynchus obliquidens	Martineau et al. (2002)	
Short-finned pilot whale	Globicephala macrorhynchus	Martineau et al. (2002)	
Harbor porpoise	Phocoena phocoena	Martineau et al. (2002)	
Amazon river dolphin	Inia geoffrensis	Martineau et al. (2002)	
Common dolphin	Delphinus delphis	Martineau et al. (2002)	
Beluga whale	Delphinapterus leucas	Martineau et al. (2002)	
Killer whale	Orcinus orca	Martineau et al. (2002)	
Pigmy sperm whale	Kogia breviceps	Martineau et al. (2002)	
Fin whale	Balaenoptera physalus	Martineau et al. (2002)	
Blue whale	Balaenoptera musculus	Martineau et al. (2002)	
Duck-billed dinosaur	Hadrosauridae	Rothschild et al. (2003)	

This beluga population, however, appears to be exceptional. Reported cancer risk in no other cetacean population approaches that of the St. Lawrence belugas. Indeed, Martineau et al. (2002) identified only 33 other cases of cancer in cetaceans worldwide prior to 2002 and note that few cancers have been found in cetaceans killed by hunters or dying of natural causes, including beluga

(continued)

in the Beaufort Sea, pilot whales, bottlenose dolphins, harbor porpoises, and various other toothed whales (Odontocetes). These researchers conclude that "cancer in stranded [St. Lawrence Estuary] belugas are more numerous than in other cetaceans, where cancer is a rare event." One may perhaps debate the latter portion of this statement on the grounds that, without careful epidemiological study directed specifically at cancer in all whales, we are largely ignorant of any measure of cancer incidence except for a few species in the order. Nevertheless, we find it interesting that in an order with the largest mammals on the planet and with species that range in mass from the vaquita (*Phocoena sinus*) (30-55 kg) to the blue whale  $(\approx 150,000 \text{ kg})$ , no correlation between cancer incidence and size has been noted.

#### Cancer in domestic mammals

Much more is known about cancer in domestic mammals than in wildlife-indeed, neoplasia is a major aspect of the pathology taught to students of veterinary medicine, including students preparing for large-animal practice and those training for care of companion animals (Hahn 2002; Withrow and Vail 2006). Also, domesticated and laboratory strain mice and rats have long been used as animal models of human cancer (Russo and Russo 1996; Hirst and Balmain 2004; Wakamatsu et al. 2007, for example). Nevertheless, we find no references in this literature reporting a correlation between cancer incidence and the product of body mass and longevity. In fact, although tumor incidence varies among small mammals (Greenacre 2004), it can reach 40% or more in "wild strain," as opposed to laboratory strain, mice (Mus musculus) (Andevort and Dunn 1962).

However, cancer incidence in domestic species yields a potentially biased view of cancer patterns among mammals in general. First, very few-less than 0.1%-of the 4000-5000 known mammalian species (Wilson 1988; Ceballos et al. 2005) have been domesticated (Watson et al. 1995; Diamond 1997, 2002). Second, many domestic animals, especially companion animals, become senescent. In humans, cancer incidence increases roughly exponentially with age usually through the first seven decades of life (Cole and Rodu 2001); therefore, cancer is primarily a disease of senescence. For example, median age at diagnosis is in the mid-60s for patients with small cell lung cancer (Van Meerbeek et al. 1997), even though this tumor type is strongly associated with environmental causes, primarily tobacco smoke (Van Meerbeek et al. 1997;

Kobzik 1999; Murren et al. 2001). Therefore, if small domestic mammals tend to live longer than do large animals, we would expect disproportionately more cancers in small mammals even under the hypothesis of equivalent cancer mechanisms among mammalian cells.

#### Peto's paradox

Under the hypothesis that cancer risk correlates with the product of body size and longevity, we would expect cancer to be common in baleen whales since they are both large and long-lived. But, as we argued earlier, the literature reveals no such pattern. Therefore, either the hypothesis is incorrect, at least for cetaceans, or we lack sufficient data to identify the pattern. The literature from other mammals suggests that the hypothesis is, in general, incorrect. Elephants, for example, are common denizens in larger zoos, and the Asian elephant (Elephas maximus), at least, has been domesticated. However, in PubMed, which indexes the major veterinary journals, a search across all fields on the terms "Elephantidae," "Elephants," "Elephas," and "Loxodonta," coupled with "tumor," "neoplasia," "cancer," and "carcinoma" in all possible permutations uncovers only 15 case reports, most of which are not cancer, and no epidemiological studies. Only two papers report tumors in Loxodonta, neither of which are cancer (a pheochromocytoma and an odontoma). A similar search for neoplasia in rhinoceroses uncovers only six case reports. In contrast, a single search under the words "Panthera AND neoplasia" finds reports of tumors in 11 lions, six tigers, two leopards, three jaguars, and one snow leopard.

Of course, we do not present these searches in lieu of epidemiological studies-we do not mean to suggest, for example, that tumor incidence in lions and elephants are roughly equal. The fact is that solid epidemiological research simply does not exist for most wildlife populations (Martineau et al. 2002), apparently including captive ones. However, suppose that cancer etiology and pathogenesis were essentially equivalent in blue whales, elephants, and roe deer. Also suppose that crude cancer risk observed for roe deer by Aguirre et al. (1999) is accurate at 2%, which is about a tenth of the risk in humans (see the beginning of this section). Roe deer weigh about 25 kg, blue whales about 150,000 kg and elephants about 4000 kg. Therefore, it takes about 6000 "roe deer equivalents" to make one blue whale and about 160 to make an elephant. Under the hypothesis of equivalent carcinogenic risk among mammalian cells,

which implies the hypothesis that randomly chosen 25 kg masses of tissue have, on average, roughly equivalent cancer risks, then the probability that a blue whale lives a cancer-free life would be less than  $(1 - 0.02)^{6000} \approx 2.27 \times 10^{-53}$ . Similarly, at least 96% of all elephants would develop cancer at some point in their lives.

Note that these calculations represent *lower* bounds for cancer risk because they are based on two intentionally incorrect assumptions. First, they assume that roe deer, blue whales and elephants all have equivalent mean life spans. In fact, the larger species live longer. Second, they assume that the number of cell divisions required to construct an organism is roughly equivalent across species, but larger organisms require more cell divisions, increasing the likelihood of oncogenic mutations. Therefore, even given the paucity of epidemiological research on wildlife neoplasia, it is somewhat surprising that so few cases of cancer in baleen whales and pachyderms have been noticed, especially since cancer has been widely reported in so many other mammals, both common and rare (Table 1).

These observations lead us to conclude that cancer risk does not correlate strongly with the product of body size and longevity; therefore, cancer etiology and pathogenesis are fundamentally different in different mammals. The reason why may seem obvious. Mammal populations vary in exposure to cancer-causing agents, cells vary in susceptibility to malignancy, cancer risk varies by tissue and mammal species vary in their proportions of each tissue type. Therefore, one should expect that cancer rates vary among mammalian species. In addition, despite striking similarities that allow us to use other mammals as models of human cancer, certain cellular mechanisms of carcinogenesis act differently in different mammalian species (Maronpot et al. 2004; Forsyth et al. 2005). However, these sources of variation could obliterate the expected correlation of cancer incidence with body mass only if they were enormous or also covaried with body mass. Therefore, just evoking variation among mammal populations-genetic, environmental, or both-does not resolve Peto's paradox.

A number of hypotheses, reviewed by Leroi et al. (2003), have been proposed that do resolve the paradox. First, mutation rates may vary inversely with body size. For example, selection may favor more effective DNA repair mechanisms in large organisms, thereby avoiding early malignant neoplasia. Second, carcinogenic mutations may be more advantageous to cells in smaller rather than larger hosts. For example, cells in larger hosts may be less

sensitive to mutations that, in smaller organisms, cause cell proliferation in preneoplastic lesions. Third, selection might favor more redundant anticancer mechanisms, like tumor-suppressor genes, within the genomes of larger organisms. To these three we may add three others. Perhaps a weak correlation between cancer incidence and body size exists, but the mechanisms of variation noted above dominate the pattern. Or, maybe there is no paradox at all. Perhaps cancer incidence covaries strongly with body size and longevity, but our general ignorance of cancer epidemiology across the class obscures the pattern. Finally, perhaps cancer case fatality correlates negatively with body size and longevity. For example, malignant neoplasia in large organisms may tend to resolve spontaneously or remain a sublethal, chronic condition, whereas the same disease would kill a smaller host. If this were true, then Peto's would be an empty paradox-the cancers are there, but we fail to notice.

In what follows, we do not attempt to identify which of these hypotheses is correct. Indeed, most are not even mutually exclusive. Instead, we adopt the more modest goal of suggesting a mechanism that may explain how cancer case fatality rate may correlate negatively with body size.

## Hypertumors and Peto's paradox

Cancer pathogenesis appears to be driven primarily by natural selection acting on phenotypic variation among neoplastic cells (Leroi et al. 2003; Nagy 2005; Axelrod et al. 2006; Merlo et al. 2006; Vineis and Berwick 2006). In the "standard model" of carcinogenesis (Cotran et al. 1999; Hanahan and Weinberg 2000), cells on the road to malignancy accumulate mutations in key genes, causing them to both proliferate and lose cohesion with the surrounding cells and with the extracellular matrix. These mutations drive cells to compete, instead of cooperate, with their genetically intact neighbors. At some point, a subpopulation of these cells acquires all the "hallmarks of cancer" (Hanahan and Weinberg 2000), including in part the ability to proliferate without external stimulation, avoid cellular senescence, shut down apoptotic mechanisms, and disperse both locally (invasion) and over long distances (metastasis). These properties give mutated cells a selective advantage as competition becomes increasingly fierce in the developing tumor. Accumulating evidence suggests that a small population of cells, called cancer stem cells, controls the behavior of most cells within a tumor (Zhang et al. 2006; O'Brien et al. 2007; Ricci-Vitiani et al. 2007).

These stem cells are not only capable of independent proliferation, but apparently can promote their own growth by directing other cells to construct an ecosystem suitable for stem cell survival and proliferation. In this sense, cancer stem cells represent the ultimate "winners" in the tumor's evolutionary game.

Angiogenesis, the growth of new blood vessels into the neoplasm, is another phenotypic property almost universally favored by selection in nascent tumors (Carmeliet and Jain 2000; Folkman et al. 2000; Hanahan and Weinberg 2000). When hypoxic, tumor cells elicit angiogenesis by secreting tumor angiogenic factors (TAF), notably vascular endothelial growth factor (VEGF) and angiopoietin-2 (ang-2) among others (Holash et al. 1998). These growth factors cause proliferation of nearby vascular endothelial cells (VECs). Daughter VECs then build new blood vessels up the VEGF concentration gradient towards the hypoxic region (Neufeld et al. 1999).

Recently, Nagy (2004, 2005) investigated in silico the consequences of natural selection and angiogenesis in malignant tumors. The core of these models consists of a system of ordinary differential equations describing the dynamics of tumor cells of various phenotypes, immature VECs able to produce mature tumoral blood vessels, and the mature vessels themselves. Competing tumor cells varied in growth potential, sensitivity to variations in local oxygen pressure affecting both birth and death rates, and ability to secrete TAF. (Model details are presented in the Supplementary Material.) Under realistic conditions, these models predict the possibility of "hypertumors," aggressive cells that fail to secrete sufficient TAF to support tumoral growth. In essence, hypertumors are composed of "cheaters" that take advantage of the vascular infrastructure built by other tumor cells. This population of grows parasitically on the tumor, cheaters damaging it perhaps to the point of inviability. Morphologically, hypertumors are expected to appear as necrotic regions associated with histological and genetic markers of aggressive growth (Nagy 2004).

This hypertumor concept may suggest a possible resolution to Peto's paradox. Larger organisms may in fact suffer more cancer, but tumors in larger organisms are more likely to evolve hypertumors, causing a negative correlation between case fatality rate and host body size. Malignant tumors in baleen whales, for example, would rarely kill their hosts because a tumor, either primary or metastatic, may experience one or more hypertumors that maintain the tumor at a sublethal size. In a pika (Ochotona princeps), on the other hand, which weighs only about 150 g (Smith and Weston 1990), most viable tumors reach lethal size before hypertumors have time to arise. Therefore, large mammals may indeed suffer more malignant neoplasia than small mammals, but the tumors would rarely represent a health risk. At most, careful necropsy of a baleen whale, for example, would reveal numerous small (maximum 10–100 g), necrotic tumors and perhaps many regression scars. In such enormous animals, however, even the largest of these tumors would be  $10^{-5}$  to  $10^{-6}$  of the animal's mass and could easily go unnoticed under conditions in which most baleen whales have been necropsied—on a whaling ship, often at sea.

#### The model and simulations

To illustrate these points and generate testable predictions, we conducted further simulations of the model described by Nagy (2004). (Simulation details are available in the online Supplementary Material.) These simulations begin with a small (0.1 g), vascularized tumor of only one cell type. At randomly chosen times, the tumor would be challenged by rare mutant cell strains. Parameters describing the phenotypes of all strains were chosen randomly from intervals representing realistic possible values. The number and mean interarrival times of mutant challengers were fixed at 20 and 50 days, respectively, although we emphasize that this choice is somewhat arbitrary. Neither parameter is well defined in existing literature. In particular, interarrival times for mutant strains cannot be equated with mutation rates because it includes the probability that a mutant strain reaches a population size large enough to be buffered from stochastic fluctuations. The values chosen, however, produce realistic results in realistic time scales.

We repeated the simulation 1000 times. In each run, we tracked tumor mass and microvessel length density (mean microvessel length per unit mass of tumor). We also monitored the number of potentially deadly tumors (defined subsequently) and time at which they would have reached this size, if at all, for six representative organisms: American pika, woodchuck, western roe deer, humans, beluga whales, and blue whales (Table 2).

Defining tumor lethality presents two main problems. First, it is well known that the distribution of tumor sizes at time of death in humans varies greatly. General host health, tumor secretory products, type and extent of involved tissues,

Table 2Estimated body and "lethal" tumor masses forrepresentative mammalian species.

Species	Body Mass (kg)	Lethal Tumor Mass (g)
Ochotona princeps	0.15	12
Marmota monax	5	170
Capreolus capreolus	25	560
Homo sapiens	70	1200
Delphinapterus leucas	1400	11,400
Balaenoptera musculus	100,000	281,000

tumor bleeding, infection, infarction, and metastatic burden all affect the size of the most massive tumor at time of death (Cotran et al. 1999). For most malignant tumors, metastatic potential determines lethality, and there is only a weak correlation between tumor size and probability of metastasis (Cotran et al. 1999). Therefore, defining a "lethal mass" for all possible cancers is impossible. One can, however, set a benchmark that represents an "average" life-threatening tumor. For humans, we set this benchmark at 1.2 kg, close to the median lethal mass for breast cancer (Spratt et al. 1993; Kuang et al. 2004). For succinctness, we will refer to this value as "lethal mass" and the frequency of tumors reaching lethal mass as "case fatality rate."

Second, it remains unclear what tumor masses produce comparable physiological effects among different-sized mammals. To our knowledge, no systematic evaluation of tumor impact exists across mammalian taxa. However, physiological and anatomical attributes, like basal metabolic rate (BMR) and brain size, are well known to scale nonlinearly with body mass, with typical allometric exponents of 2/3 to 3/4 (Wang et al. 2001; Suarez and Darveau 2005). Presumably, tumor impact would scale similarly. So, we assume, as a first approximation, that the allometric exponent for lethal tumor mass is 0.75. Using our 1.2 kg benchmark mass in humans for calibration, we obtain the following expression for tumor lethality:

### $L = 0.05 m^{0.75}$ ,

where m and L are body and lethal tumor mass, respectively, as defined earlier. Again, this relationship does not estimate actual sizes of deadly tumors; rather, it estimates the size of comparably lifethreatening tumors among mammalian species (Table 2). Also, as the allometric exponent for specific organ metabolic rates varies greatly across species (Suarez and Darveau 2005), we expect great variation in allometric scaling for specific tumor types across species. Despite these shortcomings,



**Fig. 1** Distribution of maximum tumor sizes in 1000 simulations of vascular tumor growth. In all simulations, the tumor was challenged by 20 mutant strains with a mean of 50 days between challenges.

however, the model makes practically testable predictions, as we will now describe.

#### Results

In a great majority of simulations, hypertumors held tumors to sublethal size for years, usually to the end of the simulation (Fig. 1). Only a small fraction of simulated tumors would have appeared as clinical disease in humans, with most never exceeding 10-100 g. Patterns of growth typically included early periods of rapid evolutionary change leading to more stable histology punctuated by occasional, self-limiting invasions (Fig. 2). Case fatality in the simulations correlates negatively with body mass (Fig. 3). Also, tumors take more time to reach lethal size in larger organisms (Fig. 4), but deadly tumors in larger organisms grow much faster than they do in smaller animals (Fig. 5). In conjunction with high growth rates, deadly tumors in larger organisms are also more highly vascularized (Fig. 6).

These observations are a consequence of the hypertumoral mechanism. Only tumors unencumbered by hypertumors can grow large enough to threaten the health of larger organisms. These tumors are more highly vascularized because hypertumors fail to develop, and hypertumors represent the main mechanism by which tumors in this model "outstrip their blood supply" and thereby develop ischemic necrosis.

This model generates three practically testable predictions. First, tumor necrosis will vary disproportionately with body size in mammals—tumor necrosis in small mammals should be relatively rare,



Fig. 2 Typical hypertumor dynamics. (A) Tumor mass dynamics over time. (B) Microvessel length density. Units are scaled such that 1 is the density of microvessels in the healthy tissue of tumoral origin. (C) Dynamics of individual strains. A hypertumor arises at about 300 days, which in turn is invaded by a second hypertumor at about 650 days. One can show mathematically that this tumor will eventually go extinct (data not shown).



**Fig. 3** Case fatality for six mammalian species in 1000 simulated tumors.

become more common in larger organisms, until it becomes a dominant feature of most, but not all, tumors in the largest mammals. Second, tumors that kill small mammals should vary greatly in aggression, whereas tumors that kill the largest organisms should uniformly exhibit histological and genetic markers of aggressive proliferation, including many mitotic figures in standard assays and preferential disruption of genes associated with massive tissue expansion. Finally, tumors in small mammals will also vary



Fig. 4 Time for a vascularized tumor to reach lethal mass from an initial mass of 0.1 g for six mammalian species in 1000 simulated tumors. Error bars are  $\pm 1$  SEM.



Fig. 5 Mean tumor doubling times plotted against mass of six mammalian species in 1000 simulated tumors that reached lethal mass. SEM are smaller than the diameter of the marker for each species. The line represents a least-squares regression with parameters 0.69473 (slope) and -0.97095 (intercept).

greatly in vascularization, but those that kill the largest organisms should be uniformly highly vascularized. These observations suggest a simple study compare malignant tumors in humans and whales using standard microscopic techniques measuring mitotic figures and vascular density. The St. Lawrence beluga population, for example, would be ideal for such a study. If the hypertumoral hypothesis is correct, one should observe significantly more



**Fig. 6** Mean vascularization of lethal tumors plotted against mass of the six mammalian species listed in Table 2, in 1000 simulated tumors. Error bars represent SEM.

mitotic figures, more necrosis, and higher microvessel density in the cetaceans.

#### Discussion

Although cancer has been reported in many species of mammals, most cases involve domesticated or captive animals. Very few studies allow one to estimate standard epidemiological measures like prevalence, incidence rate, lifetime risk, or case fatality for the vast majority of mammals. What little evidence there is suggests that wildlife populations suffering high incidences of cancer harbor some external etiologic agent—epizootic infection, pollution or, in the case of Tasmanian devils, a bizarre cancer able to exploit the organism's behavior.

Conspicuously absent from this pattern is a correlation with body size. Cancer does not appear to be the dominant cause of death in the largest mammals—baleen whales and elephants. Wild mice raised in captivity can suffer shockingly high rates of neoplasia (Andevort and Dunn 1962). Humans, a modestly-sized mammal, and beluga whales in the St. Lawrence, which are about 20 times more massive than humans, experience roughly similar cancer risks (Martineau et al. 2002). In contrast, cancer is rare among belugas in the Beaufort Sea (Martineau et al. 2002). These and similar observations suggest that increasing body size generally does not increase cancer risk.

The lack of correlation between body size and incidence of tumors is typically explained by differences in cellular susceptibility to oncogenic change (Leroi et al. 2003). Fewer researchers, it seems, have suggested the alternative hypothesis that malignant cells enjoy a smaller advantage in larger organisms. Hypertumors may supply a mechanistic basis that is easily testable in the field using readily available histopathologic techniques to assess tumor aggression and vascularization. To our knowledge, however, no such systematic studies have been performed.

One important aspect of malignancy ignored in the models used for this study is metastasis. Except in a few instances-for example, basal cell carcinoma or glioblastoma (Cotran et al. 1999)-metastasis characterizes cancer. It is well known that prognosis tends to correlate negatively with metastatic burden, a property on which most tumor staging systems are based. It is equally well known that metastatic potential differs among tumors of different types even in the same tissue of origin. For example, smallcell lung cancer in humans tends to metastasize more than other types of cancer, even those of the lung (Kobzik 1999). Evolutionarily, one can view metastasis as an adaptation to avoid competition for resources within a growing tumor (Nagy 2005). If ischemia becomes pronounced in a tumor, because of a hypertumor or some other mechanism, then one expects selection to favor the metastatic phenotype. Therefore, one might predict higher metastatic loads in larger mammals, like baleen whales, compared to humans. However, the simulation results suggest that hypertumors can keep most of these metastatic tumors at bay, as well. Eventually, though, the main driver of the evolutionary dynamics would switch from competition among cells to competition among tumors. Therefore, a more detailed understanding of patterns of metastasis in mammals of various sizes would be of great interest.

#### Supplementary data

Supplementary data are avilable at ICB online.

#### Acknowledgments

This research is supported in part by NSF/NIH grant DMS/NIGMS-0342388 to J.D.N.

Conflict of interest: None declared.

#### References

Acevedo-Whitehouse KA, Constantino-Casas F, Aurioles-Gamboa D, Rodriguez-Martinez HA, Godinez-Reyes CR. 1999. Hepatic carcinoma with spleen metastasis in a California sea lion from the Gulf of California. J Wildl Dis 35:565–8.

- Aguirre AA, Bröjer C, Mörner T. 1999. Descriptive epidemiology of roe deer mortality in Sweden. J Wildl Dis 35:753–62.
- Anderson WI, Johnson RC. 1988. Testicular teratoma and seminoma in a woodchuck. Vet Pathol 25:400.
- Andevort HB, Dunn TB. 1962. Occurrence of tumors in wild house mice. J Natl Cancer Inst 28:1153–63.
- Axelrod R, Axelrod DE, Pienta KJ. 2006. Evolution of cooperation among tumor cells. Proc Nat Acad Sci USA 103:13474–9.
- Bekoff M, Gese EM. 2003. Coyote (*Canis latrans*). In: Feldhammer GA, Thompson BC, Chapman JA, editors. Wild mammals of North America: biology, management and economics. Johns Hopkins: Baltimore. p 447–59.
- Bishop JM. 1989. Viruses, genes and cancer. Amer Zool 29:653–66.
- Campo MS. 1997. Bovine papilloma virus and cancer. Vet J 154:175–88.
- Carmeliet P, Jain RK. 2000. Angiogenesis in cancer and other diseases. Nature 407:249–57.
- Ceballos G, Ehrlich PR, Soberón J, Salazar I, Fay JP. 2005. Global mammal conservation: what must we manage? Science 309:603–7.
- Cole P, Rodu B. 2001. Descriptive epidemiology: cancer statistics. In: DeVita VT, Hellman S, Rosenberg SA, editors. Cancer: principles and practice of oncology. Lippencott-Raven: Philadelphia. p 228–41.
- Cotran RS, Kumar V, Collins T. 1999. Robbin's pathologic basis of disease. 6th edn. Sanders: Philadelphia.
- De Guise S, Lagacé A, Béland P. 1994. Tumors in St. Lawrence beluga whales (*Delphinapterus leucas*). Vet Pathol 31:444–9.
- De Guise S, Martineau D, Béland P, Fournier M. 1995. Possible mechanisms of action of environmental contaminants on St. Lawrence beluga whales (*Delphinapterus leucas*). Environ Health Persp 103:73–7.
- Diamond J. 1997. Guns, germs and steel: the fates of human societies. W.W. Norton: New York.
- Diamond J. 2002. Evolution, consequences and future of plant and animal domestication. Nature 418:700–7.
- Fauquier D, Gulland F, Haulena M, Spraker T. 2003. Biliary adenocarcinoma in a stranded northern elephant seal (*Mirounga angustirostris*). J Wildl Dis 39:723–6.
- Folkman J, Hahnfeldt P, Hlatky L. 2000. Cancer: looking outside the genome. Nature Rev Mol Cell Biol 1:76–9.
- Forsyth NR, Elder FFB, Shay JW, Wright WE. 2005. Lagomorphs (rabbits, pikas and hares) do not use telomere-directed replicative aging in vitro. Mech Ageing Dev 126:685–91.
- Galis F. 1999. Why do almost all mammals have seven cervical vertebrae? Developmental constraints, *Hox* genes, and cancer. J Exp Zool 285:19–26.
- Galis F, Metz JAJ. 2003. Anti-cancer selection as a source of developmental and evolutionary constraints. BioEssays 23:1035–9.

- Greenacre CB. 2004. Spontaneous tumors of small mammals. Vet Clin North Am Exot Anim Pract 7:627–51.
- Hahn KA. 2002. Veterinary oncology. 4th edn. Butterworth-Heinemann: Oxford. p 156.
- Hanahan D, Weinberg RA. 2000. The hallmarks of cancer. Cell 100:57–70.
- Hawkins CE, et al. 2006. Emerging disease and population decline of an island endemic, the Tasmanian devil *Sarcophilus harrisii*. Biol Cons 131:307–24.
- Hirayama K, Kagawa Y, Nihtani K, Taniyama H. 1999. Thyroid C-cell carcinoma with amyloid in a red fox (*Vulpes vulpes schrenchki*). Vet Pathol 36:342–4.
- Hirst GL, Balmain A. 2004. Forty years of cancer modeling in the mouse. Eur J Cancer 40:1974–80.
- Holash J, Maisonpierre PC, Compton D, Boland P, Alexander CR, Zagzag G, Yancopolous GD, Weigand SJ. 1998. Vessel cooperation, regression and growth in tumors mediated by angiopoietins and VEGF. Science 221:1994–8.
- Jardine C, Wobeser GA, Simko E. 2004. Malignant mesenchymal tumors in two white-tailed jack rabbits (*Lepus townsendii*). J Wildl Dis 40:754–8.
- Jones Y, Wise A, Maes R, Kiupel M. 2006. Peliod hepatocellular carcinoma in a domesticated ferret (*Mustela putorius furo*). J Vet Diag Invest 18:228–31.
- Kang BC, Lee YS, Lee SK. 2004. Malignant pleural mesothelioma in a woodchuck (*Marmota monax*). J Vet Med Sci 66:1617–19.
- Kang BC, Jang DD, Lee SK. 2005. Oral leiomyosarcoma in a woodchuck (*Marmota monax*). J Vet Med Sci 67:353–5.
- Keymer IF, Wells GA, Ainsworth HL. 1999. Renal neoplasia in coypus (*Myocastor coypus*). Vet J 158:144–51.
- Klaphake E, Shoieb A, Ramsay E, Schumacher J, Craig L. 2005. Renal adenocarcinoma, hepatocellular carcinoma, and pancreatic islet cell carcinoma in a binturong (*Arctictis binturong*). J Zoo Wildl Med 36:127–30.
- Kleinschmidt S, Puff C, Baumgartner W. 2006. Metastasizing oral squamous cell carcinoma in an aged pig. Vet Pathol 43:569–73.
- Kobzik L. 1999. The lung. In: Cotran RS, Kumar V, Collins T, editors. Robbin's pathologic basis of disease. 6th edn. Sanders: Philadelphia. p 697–755.
- Kuang Y, Nagy JD, Elser JJ. 2004. Biological stoichiometry of tumor dynamics: mathematical models and analysis. Disc Cont Dyn-B 4:221–40.
- Leroi AM, Koufopanou V, Burt A. 2003. Cancer selection. Nat Rev Cancer 3:226–31.
- Letcher RJ, Norstrom RJ, Muir DCG, Sandau K, Koczanski R, Michaud R, De Guise S, Béland P. 2000. Methylsulfone PCB and DDE metabolites in beluga whale (*Delphinapterus leucas*) from the St. Lawrence River estuary and western Hudson Bay. Environ Toxicol Chem 19:1378–88.
- Liu CH, Chang CH, Chin SC, Chang PH, Zhuo YX, Lee CC. 2004. Fibrosarcoma with lung and lymph node metastases in an Asian elephant (*Elephas maximus*). J Vet Diagn Invest 16:421–3.

- Maronpot RR, Flake G, Huff J. 2004. Relevance of animal carcinogenesis findings to human cancer predictions and prevention. Toxicol Pathol 32.(Suppl 1):40–8.
- Martineau D, De Guise S, Fournier M, Shugart L, Girard C, Lagacé A, Béland P. 1994. Pathology and toxicology of beluga whales from the St. Lawrence Estuary, Québec, Canada. Past, present and future. Sci Total Environ 154:201–15.
- Martineau D, Lemberger K, Dallaire A, Labelle P, Lipscomb TP, Michel P, Mikaelian I. 2002. Cancer in wildlife, a case study: Beluga from the St. Lawrence Estuary, Québec, Canada. Environ Health Persp 110:285–92.
- Mayr B, Blauensteiner J, Edlinger A, Reifinger M, Alton K, Schaffner G, Brem G. 2000. Presence of p53 mutations in feline neoplasms. Res Vet Sci 68:63–70.
- Menne S, Cote PJ. 2007. The woodchuck as an animal model for pathogenesis and therapy of chronic hepatitis B virus infection. World J Gastroenterol 13:104–24.
- Merlo LMF, Pepper JW, Reid BJ, Maley CC. 2006. Cancer as an evolutionary and ecological process. Nat Rev Cancer 6:924–35.
- Michell AR. 1999. Longevity of British breeds of dog and its relationships with sex, size, cardiovascular variables and disease. Vet Rec 145:625–9.
- Muir DCG, Ford CA, Rosenberg B, Norstrom RJ, Simon M, Béland P. 1996. Persistent organochlorines in beluga whales (*Delphinapterus leucas*) from the St. Lawrence River estuary. I. Concentrations and patterns of specific PCBs, chlorinated pesticides and polychlorinated dibenzo-p-dioxins and dibenzofurans. Environ Pollut 93:219–34.
- Muirhead CR, Kendall GM, Darby SC, Doll R, Haylock RGE, O'Hagan JA, Berridge GLC, Phillipson MA, Hunter N. 2004. Epidemiological studies of UK test veterans: II. Mortality and cancer incidence. J Radiol Prot 24:219–41.
- Murata K, Yanai T, Aqatsuma T, Uni S. 2003. *Dirofilaria immitis* infection of a snow leopard (*Uncia uncia*) in a Japanese zoo with mitochondrial DNA analysis. J Vet Med Sci 65:945–7.
- Murren J, Glatstein E, Pass HI. 2001. Small cell lung cancer. In: DeVita VT, Hellman S, Rosenberg SA, editors. Cancer: principles and practice of oncology. Lippencott-Raven: Philadelphia. p 983–1018.
- Nagy JD. 2004. Competition and natural selection in a mathematical model of cancer. Bull Math Biol 66:663–87.
- Nagy JD. 2005. The ecology and evolutionary biology of cancer: a review of mathematical models of necrosis and tumor cell diversity. Math Biosci Eng 2:381–418.
- Neufeld G, Cohen T, Gengrinovitch S, Poltorak Z. 1999. Vascular endothelial growth factor (VEGF) and its receptors. FASEB 13:9–22.
- O'Brien CA, Pollett A, Gallinger S, Dick JE. 2007. A human colon cancer cell capable of initiating tumour growth in immunodeficient mice. Nature 445:106–10.
- Paglia DE, Radcliffe RW. 2000. Anthracycline cardiotoxicity in a black rhinoceros (*Diceros bicornis*): evidence for

impaired antioxidant capacity compounded by iron overload. Vet Pathol 37:86-8.

- Palmarini M, Sharp JM, de las Heras, M Fan H. 1999. Jaagsiekte sheep retrovirus is necessary and sufficient to induce a contagious lung cancer in sheep. J Virol 73:6964–72.
- Parnell PG, Crossland JP, Beattie RM, Dewey MJ. 2005. Frequent Harderian gland adenocarcinomas in inbred white-footed mice (*Peromyscus leucopus*). Comp Med 55:382–6.
- Pearse A-M, Swift K. 2006. Transmission of devil facialtumour disease. Nature 439:549.
- Pereira MJR, Rebelo H, Teeling EC, O'Brien SJ, Mackie I, Bu SSH, Swe KM, Mie KM, Bates PJJ. 2006. Status of the world's smallest mammal, the bumble-bee bat *Craseonycteris thonglongyai* in Myanmar. Oryx 40:456–63.
- Peto R, Roe FJ, Lee PN, Levy L, Clack J. 1975. Cancer and ageing in mice and men. Br J Cancer 32:411–26.
- Plummer CE, Smith S, Andrew SE, Lassaline ME, Gelatt KN, Brooks DE, Kallberg ME, Ollivier FJ. 2007. Combined keratectomy, strontium-90 irradiation, and permanent bulbar conjunctival grafts for corneolimbal squamous cell carcinomas in horses (1990–2002): 38 horses. Vet Ophthalmol 10:37–42.
- Podell M, Pokras M, Gerlach P, Jakowski R. 1988. Meningioma in a woodchuck exhibiting central vestibular deficits. J Wildl Dis 24:695–9.
- Powe J, Castleman W, Fiorello C. 2005. A thymic carcinoid in a Bengal tiger (*Panthera tigris*). J Zoo Wildl Med 36:531–3.
- Prater MR, Duncan RB, Gaydos J. 1999. Characterization of metastatic intestinal adenocarcinoma with differentiation into multiple morphologic cell types in a Virginia opossum. Vet Pathol 36:463–8.
- Rao AT, Acharjyo LN. 1985. Squamous cell carcinoma in the lungs of a golden cat (*Felis temmincki*). J Zoo Anim Med 16:6–8.
- Raymond JT, White MR. 1999. Necropsy and histopathologic findings in 14 African hedgehogs (*Atelerix albiventris*): a retrospective study. J Zoo Wildl Med 30:273–7.
- Ricci-Vitiani L, Lombardi DG, Pilozzi E, Biffoni M, Todaro M, Peschle C, De Maria R. 2007. Identification and expansion of human colon-cancer-initiating cells. Nature 445:111–5.
- Roemer GW, Coonan TJ, Munson L, Wayne RK. 2004. Island fox (*Urocyon littoralis*). In: Sillero-Zubiri C, Hoffmann M, Macdonald WL, editors. Canids: foxes, wolves, jackals and dogs. IUCN, SSC Canid Specialist Group: Switzerland: Gland. p 97–105.
- Roth L, King JM, Hornbuckle WE, Harvey HJ, Tennant BC. 1985. Chronic hepatitis and hepatocellular carcinoma associated with persistent woodchuck hepatitis virus infection. Vet Pathol 22:338–43.
- Roth L, King JM, Tennant BC. 1991. Hepatic lesions in woodchucks (*Marmota monax*) seronegative for woodchuck hepatitis virus. J Wildl Dis 27:281–7.

- Rothschild BM, Tanke DH, Helbling M 2nd, Martin LD. 2003. Epidemiologic study of tumors in dinosaurs. Naturwissenschaften 90:495–500.
- Russo IH, Russo J. 1996. Mammary gland neoplasia in long-term rodent studies. Environ Health Perspect 104:938–67.
- Sakai H, Yanai T, Yonemaru K, Hirata A, Masiqi T. 2003. Gallbladder adenocarcinomas in two captive African lions (*Panthera leo*). J Zoo Wildl Med 34:302–6.
- Singh BP, Patterson-Kane JC, Redrobe SP, Chapman JL. 2005. Intrarenal pelvic nephroblastoma in a meerkat (*Suricata suricatta*). J Vet Diagn Invest 17:623–5.
- Smith AT, Weston ML. 1990. Ochotona princeps. J Mammal 352:1–8.
- Snyder RL, Tyler G, Summers J. 1982. Chronic hepatitis and hepatocellular carcinoma associated with woodchuck hepatitis virus. Am J Pathol 107:422–5.
- Spratt JA, von Fournier D, Spratt JS, Weber EE. 1993. Decelerating growth and human breast cancer. Cancer 71:2013–19.
- Suarez RK, Darveau CA. 2005. Multi-level regulation and metabolic scaling. J Exp Biol 208:1627–34.
- Syverton JT, Dascomb HE, Wells EB, Koomen J Jr, Berry GP. 1950. The virus-induced rabbit papilloma-to-carcinoma sequence. II. Carcinomas in the natural host, the cottontail rabbit. Cancer Res 10:440–4.
- van Meerbeek JP, et al. 1997. Life and death with small cell lung cancer (SCLC) in Flanders. Lung Cancer 18:226.
- Vineis P, Berwick M. 2006. The population dynamics of cancer: a Darwinian perspective. Int J Epidemiol 35:1151–9.
- Vitovec J. 1982. Renal cell carcinoma in a camel (*Camelus dromedarius*). Vet Pathol 19:331–3.

- Wakamatsu N, Devereux TR, Hong HH, Sills RC. 2007. Overview of the molecular carcinogenesis of mouse lung tumor models of human lung cancer. Toxicol Pathol 35:75–80.
- Wang Z, O'Connor P, Heshka S, Heymsfield SB. 2001. The reconstruction of Kleiber's law at the organ-tissue level. J Nutr 131:2967–70.
- Watson RT, Dias B, Gámez R, Heywood VH, Janetos T, Reid WV, Ruark G. 1995. Global Biodiversity Assessment: a summary for policymakers. Cambridge University Press: Cambridge.
- Whitney KM, Valentine BA, Schlafer DH. 2000. Caprine genital leiomyosarcoma. Vet Pathol 37:89–94.
- Wilson EO. 1988. The current state of biological diversity. In: Wilson EO, editor. Biodiversity. National Academy Press: Washington D.C., p 3–18.
- Withrow SJ, Vail DM. 2006. Withrow and MacEwan's small animal clinical oncology. 4th edn. Saunders: Philadelphia.
- Wooldridge AA, Gill MS, Lemarchand T, Eilts B, Taylor HW, Otterson T. 1999. Gynecomastia and mammary gland adenocarcinoma in a Nubian buck. Can Vet J 40:663–5.
- Yoon BI, Kweon OK, Kwon SW, Shin NS, Seo IB, Kim DY. 1999. Concurrent multicentric hemangiosarcoma and ovarian teratoma in an aged Pere david's deer (*Elaphurus davidianus*). J Zoo Wildl Med 30:456–8.
- Young RA, Webster WS. 1985. Tumors and polycystic renal disease in two captive woodchucks (*Marmota monax*). Lab Anim Sci 35:493–6.
- Zhang P, Zuo H, Ozaki T, Nakagomi N, Kakudo K. 2006. Cancer stem cell hypothesis in thyroid cancer. Pathol International 56:485–9.